



The path to legendary discovery™

LEGEND MAX™

ELISA Kit with Pre-coated Plates



Human IL-17F

Cat. No. 435707 1 Plate

435708 5 Plates

ELISA Kit for Accurate Quantitation of Human IL-17F
from Cell Culture Supernatant, Serum, Plasma and
Other Biological Fluids

BioLegend, Inc.
biolegend.com

It is highly recommended that this manual be read in its entirety before using this product. Do not use this kit beyond the expiration date.

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LEGEND MAX™ Human IL-17F ELISA Kit

Introduction:

IL-17F is a member of the IL-17 cytokine family which consists of at least seven structurally related proteins (IL-17A, B, C, D, E, F, and A/F). IL-17F homodimer adopts a cystine knot motif formed through the interactions of four cysteines, one of which is responsible for inter-chain bonding. IL-17F is most closely related to IL-17A, sharing 50% amino acid sequence homology. IL-17F is expressed in activated Th17 cells, along with IL-17A homodimers and IL-17A/F heterodimers. It is also expressed in activated monocytes. IL-17F has been shown to inhibit endothelial cell angiogenesis and induce increased production of IL-2, TGF- β , and MCP-1, playing a critical role in the regulation of inflammatory reactions.

The LEGEND MAX™ Human IL-17F ELISA kit is a Sandwich Enzyme-Linked Immunosorbent Assay (ELISA) with a 96-well strip plate that is pre-coated with a capture antibody. This kit is specifically designed for the accurate quantitation of human IL-17F from cell culture supernatant, serum, plasma and other biological fluids. It is analytically validated with ready-to-use reagents.

Materials Provided:

Description	Quantity (1 plate)	Quantity (5 plates)	Volume (per bottle)	Part #
Anti-Human IL-17F Pre-coated 96-well Strip Microplate	1 plate	5 plates		78581
Human IL-17F Detection Antibody	1 bottle	5 bottles	12 mL	78582
Human IL-17F Standard	1 vial	5 vials	lyophilized	78583
Matrix D (for serum and plasma samples only)	1 bottle	5 bottles	lyophilized	78339
Avidin-HRP B	1 bottle	5 bottles	12 mL	78230
Assay Buffer B	1 bottle	5 bottles	25 mL	79128
Wash Buffer (20X)	1 bottle	5 bottles	50 mL	78233
Substrate Solution F	1 bottle	5 bottles	12 mL	79132
Stop Solution	1 bottle	5 bottles	12 mL	79133
Plate Sealers	4 sheets	20 sheets		78101

Materials to be Provided by the End-User:

- Microplate reader able to measure absorbance at 450 nm
- Adjustable pipettes to measure volumes ranging from 1 µL to 1,000 µL
- Deionized water
- Wash bottle or automated microplate washer
- Log-Log graph paper or software for data analysis
- Tubes to prepare standard dilutions
- Timer
- Plate Shaker
- Polypropylene vials

Storage Information:

Store unopened kit components between 2°C and 8°C. Do not use this kit beyond its expiration date.

Opened or Reconstituted Components	
Microplate wells	If not all microplate strips are used, remove the excess strips by pressing up from underneath each strip. Place excess strips back in the foil pouch with the included desiccant pack and reseal. Store between 2°C and 8°C for up to one month.
Standard	The remaining reconstituted standard stock solution or Matrix D can be aliquoted into polypropylene vials and stored at -70°C for up to one month. Avoid repeated freeze-thaw cycles.
Matrix D	
Detection Antibody	Store opened reagents between 2°C and 8°C and use within one month.
Avidin-HRP B	
Assay Buffer B	
Wash Buffer (20X)	
Substrate Solution F	
Stop Solution	

Health Hazard Warnings:

1. Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin. Refer to the MSDS online at BioLegend’s website for details (www.biolegend.com/msds).
2. Substrate Solution F is harmful if inhaled or ingested. Avoid skin, eye and clothing contact.

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3. To reduce the likelihood of blood-borne transmission of infectious agents, handle all serum, plasma and other biological fluids in accordance with NCCLS regulations.
4. Stop Solution contains strong acid. *Wear eye, hand, and face protection.*
5. Before disposing of the plate, rinse it with an excess amount of tap water.

Specimen Collection and Handling:

Specimens should be clear and non-hemolyzed. If possible, unknown samples should be run at a number of dilutions to determine the optimal dilution factor that will ensure accurate quantitation.

Cell Culture Supernatant: If necessary, centrifuge all samples to remove debris prior to analysis. It is recommended that samples be stored at -70°C. Avoid repeated freeze-thaw cycles.

Serum: Use a serum separator tube and allow clotting for at least 30 minutes, then centrifuge for 10 minutes at 1,000 x *g*. Remove serum layer and assay immediately or store serum samples at < -70°C. Avoid repeated freeze-thaw cycles.

Plasma: Collect blood samples in a citrate, heparin or EDTA containing tube. Centrifuge for 10 minutes at 1,000 x *g* within 30 minutes of collection. Assay immediately or store plasma samples at < -70°C. Avoid repeated freeze-thaw cycles.

Reagent and Sample Preparation:

Note: All reagents should be diluted immediately prior to use.

1. Dilute the 20X Wash Buffer to 1X with deionized water. For example, make 1 liter of 1X Wash Buffer by adding 50 mL of 20X Wash Buffer to 950 mL of deionized water. If crystals have formed in the 20X Wash Buffer, bring to room temperature and vortex until dissolved.
2. Reconstitute the lyophilized Human IL-17F Standard by adding the volume of Assay Buffer B indicated on the vial label to make the 20 ng/mL standard stock solution. Allow the reconstituted standard to sit at room temperature for 15 minutes, then briefly vortex to mix completely.
3. In general, samples are analyzed without dilutions. However, if dilutions are required, use Assay Buffer B as the cell culture supernatant sample diluent and Matrix D as the serum or plasma sample diluent.

- If serum or plasma samples will be assayed, reconstitute the lyophilized Matrix D by dispensing 2 mL of deionized water into the vial and allow the reconstituted Matrix D to sit at room temperature for 15 minutes, then vortex to mix completely.

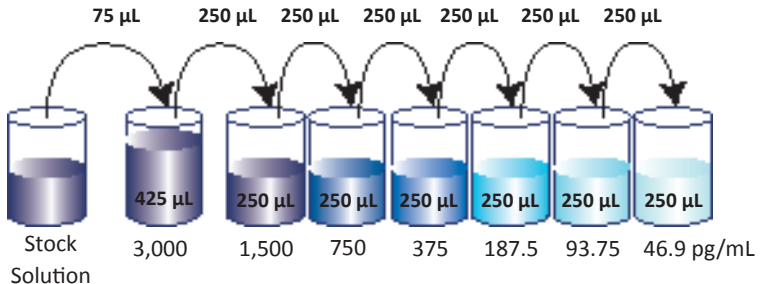
Assay Procedure:

Note: Reagents and/or antibodies from different manufacturers should not be used with this kit.

- Bring all reagents to room temperature prior to use. It is strongly recommended that all standards and samples be run in duplicate or triplicate. A standard curve is required for each assay.
- If not all microplate strips will be used, remove the excess strips by pressing up from underneath each strip. Place excess strips back in the foil pouch with the included desiccant pack and reseal.

3. For measuring samples of cell culture supernatant:

- Prepare 500 μL of the 3,000 pg/mL top standard by diluting 75 μL of the standard stock solution in 425 μL of Assay Buffer B. Perform six two-fold serial dilutions of the 3,000 pg/mL top standard in separate tubes using Assay Buffer B as the diluent. Thus, the Human IL-17F standard concentrations in the tubes are 3,000 pg/mL, 1,500 pg/mL, 750 pg/mL, 375 pg/mL, 187.5 pg/mL, 93.75 pg/mL, and 46.9 pg/mL, respectively. Assay Buffer B serves as the zero standard (0 pg/mL).

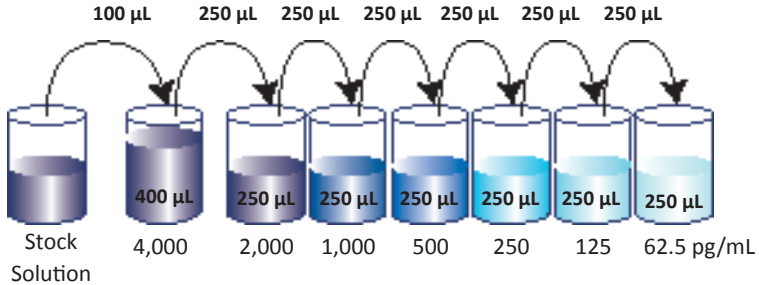


- Wash the plate 4 times with at least 300 μL of 1X Wash Buffer per well and blot any residual buffer by firmly tapping the plate upside down on absorbent paper. All subsequent washes should be performed similarly.
- Add 50 μL of Assay Buffer B to each well that will contain either standard dilutions or samples.
- Add 50 μL of standard dilutions or samples to the appropriate wells.
- Seal the plate with a Plate Sealer included in the kit and incubate the plate at room temperature for 2 hours while shaking at 200 rpm.

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4. For measuring serum or plasma samples:

- a) Prepare 500 μL of the 4,000 pg/mL top standard by diluting 100 μL of the standard stock solution in 400 μL of Assay Buffer B. Perform six two-fold serial dilutions of the 4,000 pg/mL top standard in separate tubes using Assay Buffer B as the diluent. Thus, the Human IL-17F standard concentrations in the tubes are 4,000 pg/mL , 2,000 pg/mL , 1,000 pg/mL , 500 pg/mL , 250 pg/mL , 125 pg/mL , and 62.5 pg/mL , respectively. Assay Buffer B serves as the zero standard (0 pg/mL).



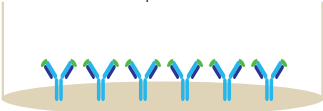
- b) Wash the plate 4 times with at least 300 μL of 1X Wash Buffer per well and blot any residual buffer by firmly tapping the plate upside down on absorbent paper. All subsequent washes should be performed similarly.
- c) Add 50 μL of Matrix D to each well that will contain standard dilutions and 50 μL of Assay Buffer B to each well that will contain samples.
- d) Add 50 μL of standard dilutions or samples to the appropriate wells.
- e) Seal the plate with a Plate Sealer included in the kit and incubate the plate **between 2°C and 8°C overnight** (16-18 hours).
5. Discard the contents of the plate into a sink, then wash the plate 4 times with 1X Wash Buffer as in step 3b or 4b.
6. Add 100 μL of Human IL-17F Detection Antibody solution to each well, seal the plate and incubate at room temperature for 1 hour while shaking.
7. Discard the contents of the plate into a sink, then wash the plate 4 times with 1X Wash Buffer as in step 3b or 4b.
8. Add 100 μL of Avidin-HRP B solution to each well, seal the plate and incubate at room temperature for 30 minutes while shaking.
9. Discard the contents of the plate into a sink, then wash the plate 5 times with 1X Wash Buffer as in step 3b and 4b. For this final wash, soak wells in 1X Wash Buffer for 30 seconds to 1 minute for each wash. This will help

minimize background.

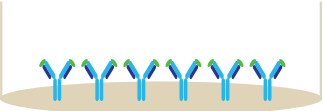
10. Add 100 μ L of Substrate Solution F to each well and incubate for 15 minutes in the dark. Wells containing human IL-17F should turn blue in color with an intensity proportional to its concentration. It is not necessary to seal the plate during this step.
11. Stop the reaction by adding 100 μ L of Stop Solution to each well. The solution color should change from blue to yellow.
12. Read absorbance at 450 nm within 30 minutes. If the reader is capable of reading at 570 nm, the absorbance at 570 nm can be subtracted from the absorbance at 450 nm.

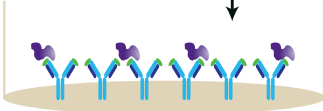
Assay Procedure Summary

- For measuring cell culture supernatant:**
Wash 4 times
Add 50 μ L Assay Buffer B to standard and sample wells

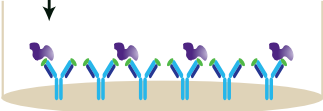


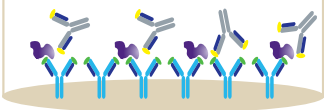
For measuring serum or plasma:
Wash 4 times
Add 50 μ L Matrix D to standard wells
Add 50 μ L Assay Buffer B to sample wells

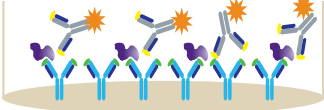

- Add 50 μ L diluted standards to standard wells
Add 50 μ L samples to sample wells
Incubate 2 hr, RT, shaking

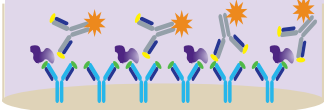


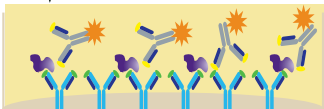
Add 50 μ L diluted standards to standard wells
Add 50 μ L samples to sample wells
Incubate **overnight, 4°C**, without shaking


- Wash 4 times
Add 100 μ L Detection Antibody solution
Incubate 1 hr, RT, shaking


- Wash 4 times
Add 100 μ L Avidin-HRP B solution
Incubate 30 min, RT, shaking


- Wash 5 times
Add 100 μ L Substrate Solution F
Incubate 15 min, RT, in the dark


- Add 100 μ L Stop Solution


- Read absorbance at 450 nm and 570 nm

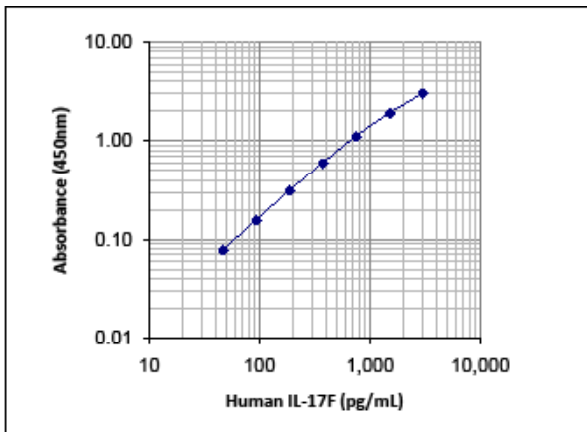
Calculation of Results:

The data can be best calculated with computer-based curve-fitting software using a 5- or 4-parameter logistics curve-fitting algorithm. If an appropriate software is not available, use log-log graph paper to determine sample concentrations. Determine the mean absorbance for each set of duplicate or triplicate standards, controls, and samples. Plot the standard curve on log-log graph paper with cytokine concentration on the X-axis and absorbance on the Y-axis. Draw a best fit line through the standard points. To determine the unknown cytokine concentrations, find the mean absorbance value of the unknown concentration on the Y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the cytokine concentration.

If samples were diluted, multiply the concentration by the appropriate dilution factor. If a test sample's absorbance value falls outside the linear portion of the standard curve, the test sample needs to be re-analyzed at a higher (or lower) dilution as appropriate.

Typical Data:

This standard curve was generated at BioLegend for demonstration purposes only. A standard curve must be run with each assay.



Performance Characteristics:

Specificity: This kit has cross-reactivity with human IL-17A/F(2.6%). No cross reactivity was observed when this kit was used to analyze the following recombinant cytokines/chemokines at up to 50 ng/mL:

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Human	IL-1 α , IL-1 β , IL-2, IL-3, IL-6, IL-8, IL-10, IL-13, IL-15, IL-17A, IL-17E, IL-22, IL-32 α , IL-33, IFN- γ , TNF- α , FGF-basic, TSLP
Mouse	IL-1 α , IL-1 β , IL-2, IL-10, IL-13, IL-22, IL-17A, IL-17A/F, IL-17B, FGF-basic, MCP-1, IFN- γ , SCF, TNF- α , GM-CSF
Rat	IL-2, IL-33, TNF- α

Sensitivity: The minimum detectable concentration is 10.7 pg/mL.

Recovery: Recombinant IL-17F (2,000, 500, and 125 pg/mL) was spiked into seven human serum samples and analyzed with the LEGEND MAX™ Human IL-17F ELISA Kit. On average, 97.7% of the cytokine was recovered from serum samples.

Linearity: Four human serum samples were spiked with high concentrations of IL-17F, then diluted with Matrix D to produce sample concentrations within the dynamic range of the assay. On average, 109.6% of the expected cytokine was detected from serum samples.

Intra-Assay Precision: Two samples containing different IL-17F concentrations were tested with sixteen replicates in one assay.

Concentration	Sample 1	Sample 2
Number of Replicates	16	16
Mean Concentration (pg/mL)	1,982.9	499.5
Standard Deviation	102.7	30.7
% CV	3.9	4.3

Inter-Assay Precision: Two samples containing different concentrations of IL-17F were tested in four independent assays.

Concentration	Sample 1	Sample 2
Number of Assays	4	4
Mean Concentration (pg/mL)	2,076.7	512.8
Standard Deviation	92.8	41.3
% CV	4.4	8.1

Biological Samples:

Serum/Plasma - Normal human serum and plasma (n = 34) were assayed for basal levels of hIL-17F. One sample was in the range of 250-500 pg/mL, eleven samples were in the range of 62.5-250 pg/mL, and twenty-two samples measured less than the lowest IL-17F standard curve point, 62.5 pg/mL.

Cell Culture Supernates - Human PBMC (1×10^6 cells/mL) were stimulated with anti-CD3 antibody for 4 days. Cell culture supernatants were collected and assayed for the levels of natural human IL-17F. The resulting IL-17F concentration was 2,562 pg/mL in anti-CD3 stimulated samples with no detectable IL-17F in unstimulated samples.

Troubleshooting Guide:

Problem	Probable Cause	Solution
High Background	Background wells were contaminated	Avoid cross-well contamination by using the provided plate sealers. Use multichannel pipettes and change tips between pipetting samples and reagents.
	Insufficient washes	Increase number of washes. Increase soaking time between washes prior to addition of substrate solution.
	TMB Substrate Solution was contaminated	TMB Substrate Solution should be clear and colorless prior to addition to wells. Use a clean container prior to pipetting substrate solution into wells.
No or poor signal	Detection Antibody, Avidin-HRP or Substrate solution were NOT added	Rerun the assay and follow the protocol.
	Wrong reagent or reagents were added in wrong sequential order	
	Insufficient plate agitation	The plate should be agitated during all incubation steps using a plate shaker at a speed where solutions in wells are within constant motion without splashing.
	The wash buffer contains Sodium Azide (NaN ₃)	Avoid Sodium Azide contamination in the wash buffer as it inhibits HRP activity.
	Incubations were done at an inappropriate temperature, timing or without agitation	Rerun the assay and follow the protocol.
Low or poor standard curve signal	The standard was incorrectly reconstituted or diluted	Adjust the calculations and follow the protocol.
	Standard was inappropriately stored	Store the reconstituted standard stock solution in polypropylene vials at -70°C. Avoid repeated freeze-thaw cycles.
	Reagents added to wells with incorrect concentrations	Check for pipetting errors and the correct reagent volume.

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Problem	Probable Cause	Solution
Signal is high, standard curves have saturated signal	Standard reconstituted with less volume than required	Reconstitute new lyophilized standard with the correct volume of solution recommended in the protocol.
	Standards/samples, detection antibody, Avidin-HRP or substrate solution were incubated for too long	Rerun the assay and follow the protocol.
Sample readings are out of range	Samples contain no or below detectable levels of the analyte	If samples are below detectable levels, it may be possible to use a larger sample volume. Contact technical support for appropriate protocol modifications.
	Samples contain analyte concentrations greater than highest standard point	Samples may require dilution and analysis.
High variation in samples and/or standards	Multichannel pipette errors	Confirm that pipette calibrations are accurate.
	Plate washing was not adequate or uniform	Ensure pipette tips are tightly secured. Ensure uniformity in all wash steps.
	Non-homogenous samples	Thoroughly mix samples before assaying.
	Samples may have high particulate matter	Remove particulate matter by centrifugation.
	Cross-well contamination	Do not reuse plate sealers. Always change tips for reagent additions. Ensure that pipette tips do not touch the reagents on the plate.

ELISA Plate Template

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												



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