

# **LEGEND MAX™**

ELISA Kit with Pre-coated Plates



# **Human Total MMP-3**

Cat. No. 444807 1 Plate

444808 5 Plates

ELISA Kit for Accurate Quantitation of Human Total MMP-3 from Cell Culture Supernatant, Serum and Heparin Plasma

BioLegend, Inc. biolegend.com

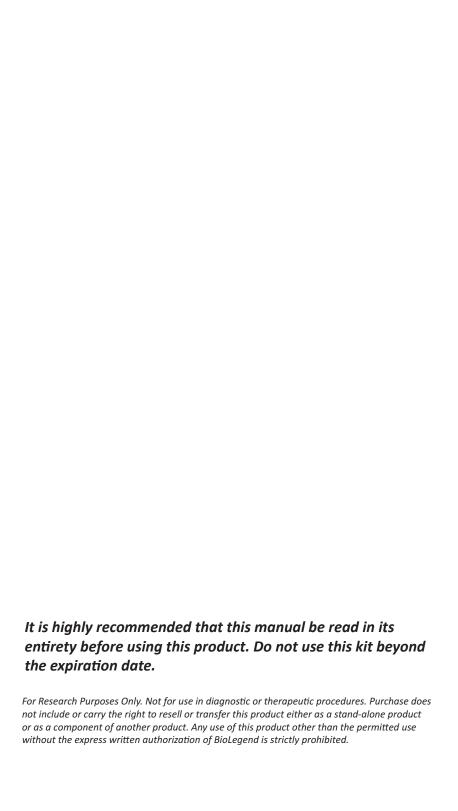


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#### Introduction:

Matrix Metalloproteinase 3 (MMP-3), also known as Matrix Metallopeptidase 3, Stromelysin 1, Progelatinase, Proteoglycanase, or Transin-1, is a member of a broad family of MMPs, which are zinc-dependent enzymes that break down extracellular matrix proteins as part of their primary functions. MMP-3 is secreted in its 54 kD inactive pro-enzyme form from many cell types including fibroblasts, chondrocytes, endothelial cells, macrophages, vascular smooth muscle cells, osteoblasts, and keratinocytes. A cysteine residue in the prodomain coordinates with the zinc in the catalytic domain to maintain latency. MMP-3 is activated by proteolytic proteins and compounds including MMP-7, 4-aminophenylmercuric acetate (APMA), and trypsin. Alpha macroglobulins and tissue inhibitor of metalloproteinase 1 (TIMP-1) are the primary natural inhibitors of MMP-3 that can regulate its activation and hinder its function. Activated MMP-3 degrades several forms of collagen, proteoglycan, fibronectin, laminin, and elastin.

Human MMP-3 shares significant amino acid sequence identity with MMP-3 from other species, including 85-99%, 83%, 81%, 77%, and 75% from non-human primates, bovine, equine, mouse, and rat, respectively.

MMP-3 is involved in tissue remodeling processes such as embryonic development, reproduction, morphogenesis, and angiogenesis. MMP-3 is a potential biomarker for many disease conditions, as it plays important roles in tumor growth, invasion, and metastasis. Additionally, MMP-3 activates several other MMP proteins by cleavage of the proenzyme domain.

The LEGEND MAX™ Human Total MMP-3 Sandwich Enzyme-Linked Immunosorbent Assay (ELISA) kit includes a 96-well strip plate that is precoated with a goat polyclonal anti-human MMP-3 antibody. The Detection Antibody is a biotinylated goat polyclonal anti-human MMP-3 antibody. This kit is specifically designed to accurately measure MMP-3, including Pro MMP-3, Mature MMP-3, and Mature MMP-3/TIMP-1 complex. This kit has been analytically validated to assess cell culture supernatant, serum, and heparin plasma samples with ready-to-use reagents.

#### **Materials Provided:**

Description	Quantity (1 plate)	Quantity (5 plates)	Volume (per bottle)	Part #
Anti-human Total MMP-3 Pre-coated 96-well Strip Microplate	1 plate	5 plates		76102
Human Total MMP-3 Detection Antibody	1 bottle	5 bottles	12 mL	76103
Human Total MMP-3 Standard	1 vial	5 vials	lyophilized	76105
Avidin-HRP	1 bottle	5 bottles	12 mL	77897
Assay Buffer A	1 bottle	5 bottles	25 mL	78232
Wash Buffer (20X)	1 bottle	5 bottles	50 mL	78233
Substrate Solution F	1 bottle	5 bottles	12 mL	79132
Stop Solution	1 bottle	5 bottles	12 mL	79133
Plate Sealers	4 sheets	20 sheets		78101

### Materials to be Provided by the End-User:

- Microplate reader able to measure absorbance at 450 nm
- Adjustable pipettes to measure volumes ranging from 1  $\mu$ L to 1,000  $\mu$ L
- · Deionized water
- Wash bottle or automated microplate washer
- Log-Log graph paper or software for data analysis
- Tubes to prepare standard dilutions
- Timer
- Plate Shaker
- Polypropylene vials

## **Storage Information:**

Store unopened kit components between 2°C and 8°C. Do not use this kit beyond its expiration date.

0	Opened or Reconstituted Components		
Microplate Strips	If not all microplate strips are used, remove the excess strips by pressing up from underneath each strip. Place excess strips back in the foil pouch with the included desiccant pack and reseal. Store between 2°C and 8°C for up to one month.		
Standard	The remaining reconstituted standard stock solution can be aliquoted into polypropylene vials and stored at -70°C for up to one month. Avoid repeated freeze-thaw cycles.		
Detection Antibody			
Avidin-HRP			
Assay Buffer A	Store opened reagents between 2°C and 8°C and use		
Wash Buffer (20X)	within one month.		
Substrate Solution F			
Stop Solution			

## **Health Hazard Warnings:**

- Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin. Refer to the MSDS online at BioLegend's website for details (www.biolegend.com/msds).
- 2. Substrate Solution F is harmful if inhaled or ingested. Avoid skin, eye and clothing contact.
- 3. To reduce the likelihood of blood-borne transmission of infectious agents, handle all serum, plasma and other biological fluids in accordance with NCCLS regulations.
- 4. Stop Solution contains strong acid and is corrosive. *Wear eye, hand, and face protection.*
- 5. Before disposing of the plate, rinse it with an excess amount of tap water.

## **Specimen Collection and Handling:**

Specimens should be clear and non-hemolyzed. If possible, unknown samples should be run at a number of dilutions to determine the optimal dilution factor that will ensure accurate quantitation.

<u>Cell Culture Supernatant</u>: If necessary, centrifuge all samples to remove debris prior to analysis. It is recommended that samples be stored at < -70°C. Avoid repeated freeze-thaw cycles.

<u>Serum</u>: Use a serum separator tube and allow clotting for at least 30 minutes, then centrifuge for 20 minutes at  $1,000 \times g$ . Remove serum layer and assay immediately or store serum samples at < -70°C. Avoid repeated freeze-thaw cycles.

<u>Plasma</u>: Collect blood samples in heparin-containing tubes, and ensure a clean stick with no hemolysis. Centrifuge immediately for 20 minutes at  $1,000 \times g$ . Carefully remove supernatant and assay immediately or store at  $<-70^{\circ}$ C. Avoid repeated freeze-thaw cycles. **EDTA and citrate are not recommended for plasma preparation due to reduced detectability of MMP-3 in the plasma.** 

#### **Reagent and Sample Preparation:**

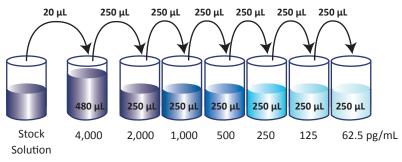
Note: All reagents should be diluted immediately prior to use.

- Dilute the 20X Wash Buffer to 1X with deionized water. For example, make 1 liter of 1X Wash Buffer by adding 50 mL of 20X Wash Buffer to 950 mL of deionized water. If crystals have formed in the 20X Wash Buffer, bring to room temperature and vortex until dissolved.
- 2. Reconstitute the lyophilized Human Total MMP-3 Standard by adding the volume of Assay Buffer A to make a 100 ng/mL standard stock solution (refer to LEGEND MAX™ Kit Lot-Specific Certificate of Analysis/LEGEND MAX™ Kit Protocol). Allow the reconstituted standard to sit at room temperature for 15 minutes, then briefly vortex to mix completely.
- 3. For measuring serum or plasma samples, a 10-fold dilution is recommended using Assay Buffer A. For example, dilute 30  $\mu L$  of sample in 270  $\mu L$  of Assay Buffer A. For cell culture supernatant samples, in general, no sample dilution is required. If dilutions are necessary, samples should be diluted with Assay Buffer A.

### **Assay Procedure:**

Note: Do not mix reagents from different kits or lots. Reagents and/or antibodies from different manufacturers should not be used with this kit.

- Bring all reagents to room temperature prior to use. It is strongly recommended that all standards and samples be run in duplicate or triplicate. A standard curve is required for each assay.
- 2. If not all microplate strips will be used, remove the excess strips by pressing up from underneath each strip. Place excess strips back in the foil pouch with the included desiccant pack and reseal.
- 3. Prepare 500 μL of the 4,000 pg/mL top standard by diluting 20 μL of the standard stock solution in 480 μL of Assay Buffer A. Perform six two-fold serial dilutions of the 4,000 pg/mL top standard in separate tubes using Assay Buffer A as the diluent. Thus, the human Total MMP-3 standard concentrations in the tubes are 4,000 pg/mL, 2,000 pg/mL, 1,000 pg/mL, 500 pg/mL, 250 pg/mL, 125 pg/mL, 62.5 pg/mL, respectively. Assay Buffer A serves as the zero standard (0 pg/mL).



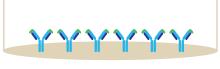
- 4. Wash the plate 4 times with 1X Wash Buffer. Wash the plate with at least 300  $\mu$ L of 1X Wash Buffer per well and blot any residual buffer by firmly tapping plate upside down on absorbent paper. All subsequent washes should be performed similarly.
- 5. Add 50  $\mu$ L Assay Buffer A to each well that will contain either standards or samples.
- 6. Add 50  $\mu$ L of standard dilutions or properly diluted samples to the appropriate wells.
- 7. Seal the plate with a Plate Sealer included in the kit and incubate the plate at room temperature for 2 hours while shaking at 200 rpm.
- 8. Discard the contents of the plate into a sink, then wash the plate 4 times with 1X Wash Buffer as in step 4.

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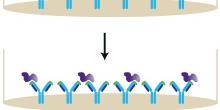
- 9. Add 100  $\mu$ L of Human Total MMP-3 Detection Antibody solution to each well, seal the plate and incubate at room temperature for 1 hour while shaking.
- 10. Discard the contents of the plate into a sink, then wash the plate 4 times with 1X Wash Buffer as in step 4.
- 11. Add 100  $\mu$ L of Avidin-HRP solution to each well, seal the plate and incubate at room temperature for 30 minutes while shaking.
- 12. Discard the contents of the plate into a sink, then wash the plate 5 times with 1X Wash Buffer as in step 4. For this final wash, soak wells in 1X Wash Buffer for 30 seconds to 1 minute for each wash. This will help minimize background.
- 13. Add 100  $\mu$ L of Substrate Solution F to each well and incubate at room temperature for 15 minutes in the dark. Wells containing human Total MMP-3 should turn blue in color with an intensity proportional to its concentration. It is not necessary to seal the plate during this step.
- 14. Stop the reaction by adding 100  $\mu$ L of Stop Solution to each well. The solution color should change from blue to yellow.
- 15. Read absorbance at 450 nm immediately. If the reader is capable of reading at 570 nm, the absorbance at 570 nm can be subtracted from the absorbance at 450 nm.

## **Assay Procedure Summary**

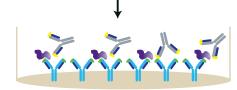
1. Wash 4 times Add 50 μL Assay Buffer A



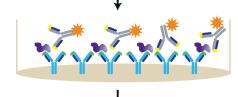
2. Add 50 µL diluted standards or samples Incubate 2 hrs, RT, shaking



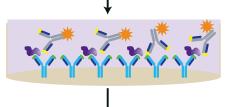
3. Wash 4 times Add 100 μL Detection Antibody solution Incubate 1 hr, RT, shaking



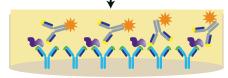
4. Wash 4 times Add 100 μL Avidin-HRP solution Incubate 30 mins, RT, shaking



5. Wash 5 times Add 100 μL Substrate Solution F Incubate 15 mins, RT, in the dark



6. Add 100 μL Stop Solution



7. Read absorbance at 450 nm and 570 nm

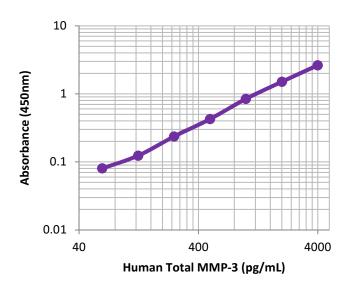
#### Calculation of Results:

The data can be best calculated with computer-based curve-fitting software using a 5- or 4-parameter logistics curve-fitting algorithm. If an appropriate software is not available, use log-log graph paper to determine sample concentrations. Determine the mean absorbance for each set of duplicate or triplicate standards, controls, and samples. Plot the standard curve on log-log graph paper with protein concentration on the X-axis and absorbance on the Y-axis. Draw a best fit line through the standard points. To determine the unknown protein concentrations, find the mean absorbance value of the unknown concentration on the Y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the protein concentration.

If samples were diluted, multiply the concentration by the appropriate dilution factor. If a test sample's absorbance value falls outside the linear portion of the standard curve, the test sample needs to be re-analyzed at a higher (or lower) dilution as appropriate.

## **Typical Data:**

This standard curve was generated at BioLegend for demonstration purposes only. A standard curve must be run with each assay.



#### **Performance Characteristics:**

<u>Specificity:</u> No cross-reactivity was observed when this kit was used to analyze the following recombinant proteins, each at 100 ng/mL except where the concentration is specified.

	MMP-2, MMP-8, MMP-9, MMP-15, MMP-16, TIMP-1, TIMP-2, CCL1 (50 ng/mL), IFN-γ (50 ng/mL), NGAL
Mouse	MMP-2, MMP-9, NGAL

This kit cross-reacts with the following proteins, tested at 100 ng/mL.

Protein	Cross-Reactivity	Protein	Cross-Reactivity
Human MMP-1	0.20%	Human MMP-12	0.22%
Human MMP-7	0.20%	Human MMP-13	0.24%
Human MMP-10	0.18%	Human MMP-14	0.41%
Human MMP-11	0.17%	Mouse MMP-3	94.02%

This kit detects human APMA-matured MMP-3 and APMA-matured MMP-3/TIMP-1 complex at ~31% signal strength relative to the Human Total MMP-3 Standard at 100 ng/mL. There is small or negligible interference caused by TIMP-1 or alpha-2-Macroglobulin to the reactivity of this kit at 122.2 ng/mL and 671.3 ng/mL at molar ratios with MMP-3 of 10:1 and 2:1, respectively.

<u>Sensitivity:</u> The minimum detectable concentration is <21.6 pg/mL  $\pm$  18.3 pg/mL (mean  $\pm$  2SD; n=11).

<u>Linearity:</u> Human samples were diluted with Assay Buffer A to produce samples with values within the dynamic range, and then assayed with the kit to determine the dilutional linearity up to 8-fold dilutions.

Sample Type	N	% Linearity
Serum	5	104
Heparin Plasma	5	108
HepG2 cell culture supernatant	1	94
U87-MG cell culture supernatant	1	99

<u>Recovery:</u> Recombinant human Total MMP-3, at concentrations of 1,000, and 250 pg/mL, was spiked into the following human samples, with the exception of heparin plasma, into which recombinant human APMA-matured MMP-3/TIMP-1 complex was spiked. \*Samples were diluted prior to spiking.

Sample Type	N	% Recovery
Serum*	5	111
Heparin Plasma*	4	99
HepG2 cell culture supernatant	1	106
U87-MG cell culture supernatant*	1	108

<u>Intra-Assay Precision:</u> Two samples with different concentrations of human heparin plasma were tested with 16 replicates in one assay.

Concentration	Sample 1	Sample 2
Number of Replicates	16	16
Mean Concentration (pg/mL)	1446	336
Standard Deviation	71	13
% CV	4.9	3.9

<u>Inter-Assay Precision:</u> Two different concentrations of human heparin plasma were assayed in four independent assays by four different users.

Concentration	Sample 1	Sample 2
Number of Assays	4	4
Mean Concentration (pg/mL)	1322	328
Standard Deviation	106	26
% CV	8.0	7.9

<u>Biological Samples:</u> Matched serum and plasma samples from 17 donors were tested in one assay.

Sample Type	N	% Detectable	Min. (pg/mL)	Max. (pg/mL)	Median (pg/mL)
Serum	17	100	2,240	30,090	10,360
Heparin Plasma	17	100	6,700	39,450	13,820

Human U87-MG, HepG2, and HEK 293 cells were established in 10% FBS/ EMEM at a concentration of  $1x10^6$ /mL and grown for a period of 4 days. Supernatant from the culture was collected on day 4, and the concentrations of Total MMP-3 in the U87-MG, HepG2, and HEK 293 cell supernatant were measured to be 80,480 pg/mL, 376 pg/mL, and 104 pg/mL, respectively.

# **Troubleshooting Guide:**

Problem	Probable Cause	Solution
High Background	Background wells were contaminated	Avoid cross-well contamination by using the provided plate sealers.
		Use multichannel pipettes and change tips between pipetting samples and reagents.
	Insufficient washes	Increase number of washes. Increase soaking time between washes prior to addition of substrate solution.
	TMB Substrate Solution was contaminated	TMB Substrate Solution should be clear and colorless prior to addition to wells. Use a clean container prior to pipetting substrate solution into wells.
No or poor signal	Detection Antibody, Avidin-HRP or Substrate solution were NOT added	Down the access and follow the protocol
	Wrong reagent or reagents were added in wrong sequential order	Rerun the assay and follow the protocol.
	Insufficient plate agitation	The plate should be agitated during all incubation steps using a plate shaker at a speed where solutions in wells are within constant motion without splashing.
	The wash buffer contains Sodium Azide (NaN3)	Avoid Sodium Azide contamination in the wash buffer as it inhibits HRP activity.
	Incubations were done at an inappropriate temperature, timing or without agitation	Rerun the assay and follow the protocol.
Low or poor standard curve	The standard was incorrectly reconstituted or diluted	Adjust the calculations and follow the protocol.
signal	Standard was inappropriately stored	Store the reconstituted standard stock solution in polypropylene vials at -70°C. Avoid repeated freeze-thaw cycles.
	Reagents added to wells with incorrect concentrations	Check for pipetting errors and the correct reagent volume.

Problem	Probable Cause	Solution
Signal is high, standard curves have saturated	Standard reconstituted with less volume than required	Reconstitute new lyophilized standard with the correct volume of solution recommended in the protocol.
signal	Standards/samples, detection antibody, Avidin-HRP or substrate solution were incubated for too long	Rerun the assay and follow the protocol.
Sample readings	Samples contain no or below detectable levels of the analyte	If samples are below detectable levels, it may be possible to use a larger sample volume. Contact technical support for appropriate protocol modifications.
are out of range	Samples contain analyte concentrations greater than highest standard point	Samples may require dilution and analysis.
	Multichannel pipette errors	Confirm that pipette calibrations are accurate.
High variation in samples and/or	Plate washing was not adequate or uniform	Ensure pipette tips are tightly secured.  Ensure uniformity in all wash steps.
standards	Non-homogenous samples	Thoroughly mix samples before assaying.
	Samples may have high particulate matter	Remove particulate matter by centrifugation.
	Cross-well contamination	Do not reuse plate sealers.
		Always change tips for reagent additions. Ensure that pipette tips do not touch the reagents on the plate.

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# **Notes**

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# **Notes**




LEGEND MAX™ Kits are manufactured by **BioLegend Inc.** 

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