BioLegend®

Enabling Legendary Discovery™

LEGEND MAX™

ELISA Kit with Pre-coated Plate

Human IFN-α2

Cat. No. 446407

ELISA Kit for Accurate Quantitation of Human IFN-α2 from Cell Culture Supernatant, Serum, Plasma and Other Biological Fluids

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biolegend.com

It is highly recommended that this manual be read in its entirety before using this product. Do not use this kit beyond the expiration date.

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Introduction:

Interferon alpha-2 (IFN-α2) is a cytokine encoded by the IFNA2 gene located on chromosome 9. It belongs to the Type 1 IFN family which includes 13 α subtypes, 1 β, and 1 ω. IFN-α2 was the first of these subtypes to be produced pharmaceutically and is the most widely studied. The mature secreted protein consists of 165 amino acids with 5 α-helices and two disulfide bonds. There are a several allelic variants of IFN-α2, including IFN-α2a, IFN-α2b, and IFN-α2c. IFN-α2 can act on all cell types due to the ubiquitous expression of the Type 1 IFN receptor (IFNAR).

IFN-α2 acts to combat infection by stimulating the production of proteins that inhibit viral replication and prevent viral entry into neighboring cells. IFN-α2 can stimulate the immune system by initiating T and NK cell mediated cytotoxicity. IFN-α2 also can inhibit cancer cell proliferation and activate immune system to target cancer cells. IFN-α2 has been used therapeutically in a variety of viral infections (including hepatitis B and C) and certain cancers (such as melanoma, renal cell carcinoma).

BioLegend's LEGEND MAX™ Human IFN-α2 ELISA kit is a Sandwich Enzyme-Linked Immunosorbent Assay (ELISA) with a 96-well strip plate that is pre-coated with a monoclonal mouse anti-human IFN-α2 capture antibody. The detection antibody is a biotinylated monoclonal mouse anti-human IFN-α2 antibody. This kit is specifically designed for the accurate quantitation of human IFN-α2 from cell culture supernatant, serum, plasma and other biological fluids. Different IFN-α2 allelic variants can be detected equally. It is analytically validated with ready-to-use reagents.

Materials Provided:

Description	Quantity	Volume	Part #
Anti-Human IFN-α2 Pre-coated 96-well Strip Microplate	1 plate		750000222
Human IFN-α2 Detection Antibody	1 bottle	12 mL	750000146
Human IFN-α2 Standard	1 vial	lyophilized	750000149
Matrix A	1 vial	lyophilized	78303
Avidin-HRP	1 bottle	12 mL	77897
Assay Buffer G	1 bottle	60 mL	750000253
Wash Buffer (20X)	1 bottle	50 mL	78233
Substrate Solution F	1 bottle	12 mL	76335
Stop Solution	1 bottle	12 mL	79133
Plate Sealers	4 sheets		78101

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Materials to be Provided by the End-User:

- Microplate reader able to measure absorbance at 450 nm
- Adjustable pipettes to measure volumes ranging from 1 µL to 1,000 µL
- Deionized water
- Wash bottle or automated microplate washer
- Log-Log graph paper or software for data analysis
- Polypropylene tubes to prepare standard dilutions
- Timer
- Plate Shaker
- Polypropylene vials

Storage Information:

Store unopened kit components between 2°C and 8°C. Do not use this kit beyond its expiration date.

<table>
<tr><th colspan="2">Opened or Reconstituted Components</th></tr>
<tr><td>Microplate wells</td><td>If all microplate strips will not be used, remove the excess strips by pressing up from underneath each strip. Place excess strips back in the foil pouch with the included desiccant pack and reseal. Store between 2°C and 8°C for up to one month.</td></tr>
<tr><td>Standard</td><td rowspan="2">The remaining reconstituted standard stock solution can be aliquoted into polypropylene vials and stored at -70°C for up to one month. Avoid repeated freeze-thaw cycles.</td></tr>
<tr><td>Matrix A</td></tr>
<tr><td>Detection Antibody</td><td rowspan="6">Store opened reagents between 2°C and 8°C and use within one month.</td></tr>
<tr><td>Avidin-HRP</td></tr>
<tr><td>Assay Buffer G</td></tr>
<tr><td>Wash Buffer (20X)</td></tr>
<tr><td>Substrate Solution F</td></tr>
<tr><td>Stop Solution</td></tr>
</table>

Health Hazard Warnings:

1. Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin. Refer to the MSDS online at BioLegend's website for details (www.biolegend.com/msds).
2. Substrate Solution F is harmful if inhaled or ingested. Avoid skin, eye and clothing contact.

3. To reduce the likelihood of blood-borne transmission of infectious agents, handle all serum, plasma and other biological fluids in accordance with NCCLS regulations.
4. Stop Solution contains strong acid. *Wear eye, hand, and face protection.*
5. Before disposing of the plate, rinse it with an excess amount of tap water.

Specimen Collection and Handling:

Specimens should be clear and non-hemolyzed. If possible, unknown samples should be run at a number of dilutions to determine the optimal dilution factor that will ensure accurate quantitation.

Cell Culture Supernatant: If necessary, centrifuge all samples to remove debris prior to analysis. It is recommended that samples be stored at < -70°C. Avoid repeated freeze-thaw cycles.

Serum: Use a serum separator tube and allow clotting for at least 30 minutes, then centrifuge for 10 minutes at 1,000 x *g*. Remove serum layer and assay immediately or store serum samples at < -70°C. Avoid repeated freeze-thaw cycles.

Plasma: Collect blood samples in citrate, heparin or EDTA containing tubes. Centrifuge for 10 minutes at 1,000 x *g* within 30 minutes of collection. Assay immediately or store plasma samples at < -70°C. Avoid repeated freeze-thaw cycles.

Reagent and Sample Preparation:

Note: All reagents should be diluted immediately prior to use.

1. Dilute the 20X Wash Buffer to 1X with deionized water. For example, make 1 liter of 1X Wash Buffer by adding 50 mL of 20X Wash Buffer to 950 mL of deionized water. If crystals have formed in the 20X Wash Buffer, bring to room temperature and vortex until dissolved.
2. Reconstitute the lyophilized Human IFN-α2 Standard by adding the volume of Assay Buffer G to make the 20 ng/mL standard stock solution (Refer to LEGEND MAX Kit Lot-Specific Certificate of Analysis/LEGEND MAX Kit Protocol). Allow the reconstituted standard to sit at room temperature for 15-20 minutes, then briefly vortex to mix completely.
3. If serum and plasma samples are to be read, reconstitute Matrix A in 2 mL of deionized water for 15-20 minutes, then briefly vortex to mix completely. If only cell culture samples are to be read, Matrix A is not required.

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4. In general, serum samples require a 2-fold dilution and plasma samples require a 4-fold dilution. Serum and Heparin Plasma are recommended, but Citrate and EDTA plasma can be used. Tissue culture supernatants do not require dilution. If dilution is required, use Assay Buffer G. Lower or higher dilutions can be used as needed, but must be quantified to fit the assay range by the end user.

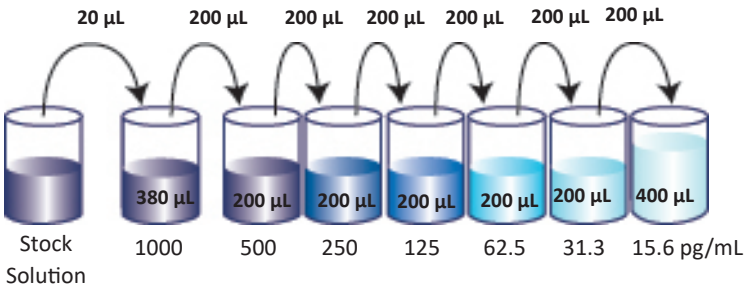
Assay Procedure:

Note: Do not mix reagents from different kits or lots. Reagents and/or antibodies from different manufacturers should not be used with this kit.

1. Bring all reagents to room temperature prior to use. It is strongly recommended that all standards and samples be run in duplicate or triplicate. A standard curve is required for each assay.

2. If all microplate strips will not be used, remove the excess strips by pressing up from underneath each strip. Place excess strips back in the foil pouch with the included desiccant pack and reseal.

3. Add 20 µL of the 20 ng/mL stock solution to 380 µL of Assay Buffer G in a separate test tube to generate the top standard. Perform six two-fold serial dilutions of the 20 ng/mL top standard using Assay Buffer G as the diluent. Thus, the Human IFN-α2 standard concentrations in the tubes are 1000 pg/mL, 500 pg/mL, 250 pg/mL, 125 pg/mL, 62.5 pg/mL, 31.3 pg/mL and 15.6 pg/mL, respectively. Assay Buffer G serves as the zero standard (0 pg/mL).



4. Wash the plate 4 times with at least 300 µL of 1X Wash Buffer per well and blot any residual buffer by firmly tapping the plate upside down on absorbent paper. All subsequent washes should be performed similarly

5. For Cell Culture samples, add 50 µL of Assay Buffer G to wells that will contain standard dilutions. For serum or plasma samples, added 50 µL of Matrix A to each well that will contain standard dilutions. Add 50 µL of Assay Buffer G to all wells that will contain samples. Then add 50 µL of standard dilutions or samples to the appropriate wells.

6. Seal the plate with a Plate Sealer included in the kit and incubate the plate for 2 hours at room temperature while shaking.
7. Discard the contents of the plate into a sink, then wash the plate 4 times with 1X Wash Buffer as in step 4.
8. Add 100 µL of Human IFN-α2 Detection Antibody solution to each well. Seal the plate and incubate at room temperature for 1 hour while shaking.
9. Discard the contents of the plate into a sink, then wash the plate 4 times with 1X Wash Buffer as in step 4.
10. Add 100 µL of Avidin-HRP solution to each well, seal the plate and incubate at room temperature for 30 minutes while shaking.
11. Discard the contents of the plate into a sink, then wash the plate 5 times with 1X Wash Buffer as in step 4. For this final wash, soak wells in 1X Wash Buffer for 30 seconds to 1 minute for each wash. This will help minimize background.
12. Add 100 µL of Substrate Solution F to each well and incubate for 15 minutes in the dark. Wells containing human IFN-α2 should turn blue in color with intensity proportional to its concentration. It is not necessary to seal the plate during this step.
13. Stop the reaction by adding 100 µL of Stop Solution to each well. The solution color should change from blue to yellow.
14. Read absorbance at 450 nm within 30 minutes. If the reader is capable of reading at 570 nm, the absorbance at 570 nm can be subtracted from the absorbance at 450 nm.

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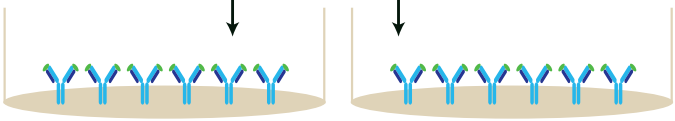
Assay Procedure Summary

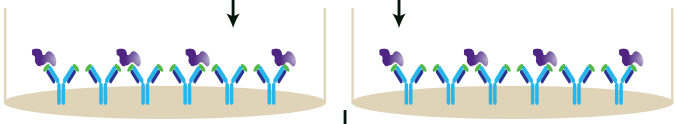
1. **For measuring cell culture supernatant:**
Add 50 µl Assay Buffer G to standard and sample wells

For measuring serum or plasma:
Add 50 µl Matrix A to standard wells
Add 50 µl Assay Buffer G to sample wells

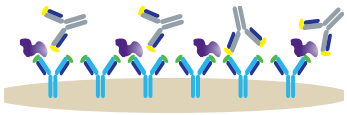


2. Add 50 µl diluted standards to standard wells
Add 50 µl samples to sample wells
Incubate 2 hr, RT, shaking

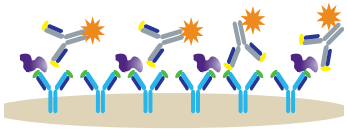
Add 50 µl diluted standards to standard wells
Add 50 µl samples to sample wells
Incubate 2 hr, RT, shaking



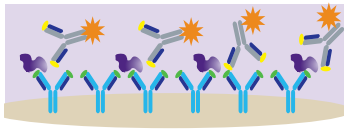
3. Wash 4 times
Add 100 µl Detection Antibody solution
Incubate 1 hr, RT, shaking



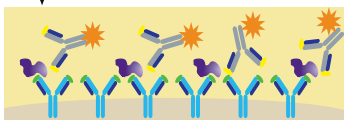
4. Wash 4 times
Add 100 µl Avidin-HRP solution
Incubate 30 min, RT, shaking



5. Wash 5 times
Add 100 µl Substrate Solution F
Incubate 15 min, RT, in the dark



6. Add 100 µl Stop Solution



7. Read absorbance at 450nm and 570nm

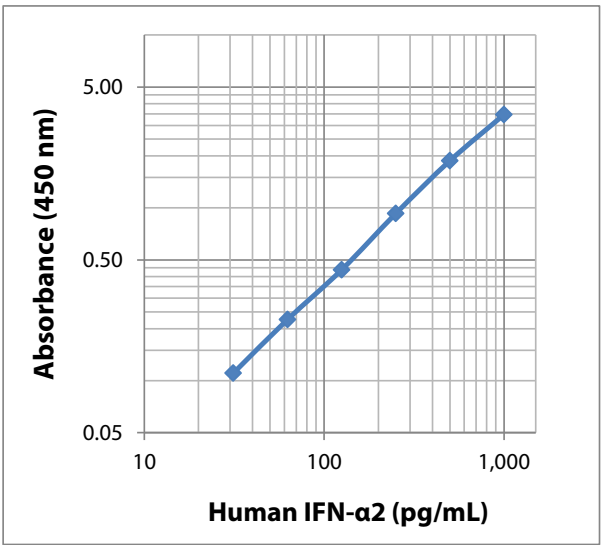
Calculation of Results:

The data can be best calculated with computer-based curve-fitting software using a 5- or 4-parameter logistics curve-fitting algorithm. If an appropriate software is not available, use log-log graph paper to determine sample concentrations. Determine the mean absorbance for each set of duplicate or triplicate standards, controls, and samples. Plot the standard curve on log-log graph paper with cytokine concentration on the X-axis and absorbance on the Y-axis. Draw a best fit line through the standard points. To determine the unknown cytokine concentrations, find the mean absorbance value of the unknown concentration on the Y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the cytokine concentration.

If samples were diluted, multiply the concentration by the appropriate dilution factor. If a test sample's absorbance value falls outside the linear portion of the standard curve, the test sample needs to be re-analyzed at a higher (or lower) dilution as appropriate.

Typical Data:

This standard curve was generated at BioLegend for demonstration purposes only. A standard curve must be run with each assay.



Performance Characteristics:

Specificity: This kit recognizes natural and recombinant human IFN-α2. No cross reactivity was observed when this kit was used to analyze the following recombinant proteins at 50 ng/mL.

Human	IFN-α1, IFN-α5, IFN-β, IFN-λ1, IFN-λ2, IFN-ω, CCL1, CCL2, CCL5, CCL8, CCL11, CCL13, CCL14, CCL17, CCL19, CCL22, CXCL1, CXCL2, CXCL3, CXCL5, CXCL12, IL-1a, IL-2, IL-8, IL-36, EPO, GM-CSF, IGF-1, Paxillin, PLGF-1, PDGF-BB, LT-a, M-CSF, NGAL, MIF, MMP-1, MMP-3, MMP-9, TGF-a, TGF-b2, TNFa, TSLP, SCF, VEGF-165, Cystatin C, ICAM-1-Fc, Resistin
Mouse	IFN-α1, IFN-β1, IFN-γ, IL-4, IL-10, IL-11, IL-12, IL-13, IL-15, IL-17, IL-33, IL-34

Kit has equal recognition between the IFN-α2 allelic variants IFN-α2a, IFN-α2b, and IFN-α2c. 100% cross reactivity with IFN-α6.

Sensitivity: The minimum detectable concentration of IFN-α2 is 2.94±0.89 pg/mL with assay buffer (n=9), and 2.57±0.64 pg/mL with Matrix A (n=10). The sensitivities shown are ± one standard deviation.

Recovery: Recombinant human IFN-α2 (1000, 250, 62.5 pg/mL) was spiked into human samples, then analyzed with the LEGEND MAX™ Human IFN-α2 ELISA kit.

Sample Type	N	% Recovery
Serum	5	87.9
Citrate Plasma	5	80.8
EDTA Plasma	5	74.2
Heparin Plasma	5	93.3

Linearity: Human samples were diluted 2-fold and then spiked with IFN-α2 recombinant protein. These were then diluted to produce samples within the dynamic range of the kit. Samples were then assayed to determine the dilutional linearity.

Sample Type	N	% Linearity
Serum	5	104.3
Citrate Plasma	5	103.3
EDTA Plasma	5	100.7
Heparin Plasma	5	104.2

Intra-Assay Precision: Two samples containing different human IFN-α2 concentrations were tested on one plate with 16 replicates.

Concentration	Sample 1	Sample 2
Number of Replicates	16	16
Mean Concentration (pg/mL)	261.3	49.0
Standard Deviation	26.9	4.1
%CV	10.3	8.3

Inter-Assay Precision: Two samples containing different human IFN-α2 concentrations were tested in four independent assays.

Concentration	Sample 1	Sample 2
Number of Assays	4	4
Mean Concentration (pg/mL)	315.4	48.9
Standard Deviation	26.7	4.3
%CV	8.5	8.8

Normal Biological Samples:

Human serum and plasma samples (n = 32) were assayed for natural human IFN-α2. Serum and plasma range detected was ND – 0.031 pg/mL. Significant detection was found in the supernatant from PBMCs stimulated with Poly I:C (176.2 pg/mL) and R848 (2884.9 pg/mL).

Troubleshooting Guide:

Problem	Probable Cause	Solution
High Background	Background wells were contaminated	Avoid cross-well contamination by using the provided plate sealers. Use multichannel pipettes and change tips between pipetting samples and reagents.
	Insufficient washes	Increase number of washes. Increase soaking time between washes prior to addition of substrate solution.
	TMB Substrate Solution was contaminated	TMB Substrate Solution should be clear and colorless prior to addition to wells. Use a clean container prior to pipetting substrate solution into wells.
No or poor signal	Detection Antibody, Avidin-HRP or Substrate solution were NOT added	Rerun the assay and follow the protocol.
	Wrong reagent or reagents were added in wrong sequential order	
	Insufficient plate agitation	The plate should be agitated during all incubation steps using a plate shaker at a speed where solutions in wells are within constant motion without splashing.
	The wash buffer contains Sodium Azide (NaN3)	Avoid Sodium Azide contamination in the wash buffer as it inhibits HRP activity.
	Incubations were done at an inappropriate temperature, timing or without agitation	Rerun the assay and follow the protocol.
Low or poor standard curve signal	The standard was incorrectly reconstituted or diluted	Adjust the calculations and follow the protocol.
	Standard was inappropriately stored	Store the reconstituted standard stock solution in polypropylene vials at -70°C. Avoid repeated freeze-thaw cycles.
	Reagents added to wells with incorrect concentrations	Check for pipetting errors and the correct reagent volume.

Problem	Probable Cause	Solution
Signal is high, standard curves have saturated signal	Standard reconstituted with less volume than required	Reconstitute new lyophilized standard with the correct volume of solution recommended in the protocol.
	Standards/samples, detection antibody, Avidin-HRP or substrate solution were incubated for too long	Rerun the assay and follow the protocol.
Sample readings are out of range	Samples contain no or below detectable levels of the analyte	If samples are below detectable levels, it may be possible to use a larger sample volume. Contact technical support for appropriate protocol modifications.
	Samples contain analyte concentrations greater than highest standard point	Samples may require dilution and analysis.
High variation in samples and/or standards	Multichannel pipette errors	Confirm that pipette calibrations are accurate.
	Plate washing was not adequate or uniform	Ensure pipette tips are tightly secured. Ensure uniformity in all wash steps.
	Non-homogenous samples	Thoroughly mix samples before assaying.
	Samples may have high particulate matter	Remove particulate matter by centrifugation.
	Cross-well contamination	Do not reuse plate sealers. Always change tips for reagent additions. Ensure that pipette tips do not touch the reagents on the plate.

ELISA Plate Template

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

LEGEND MAX™ Kits are manufactured by **BioLegend Inc.**
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